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## **Screening and confirmation of ritalinic acid in urine by LC-MS<sup>n</sup>**

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### **Introduction**

Methylphenidate (MPH) is therapeutically used for the treatment of attention-deficit/hyperactivity disorder (ADHD). In sports it can be abused to overcome fatigue or to increase the tolerance to exhaustion. Less than 1% of the administered dose is excreted unchanged in urine whereas 80% is excreted as ritalinic acid (RA) [1]. Moreover RA can be detected much longer in urine than MPH. Hence RA is the preferred target compound to detect MPH administration [1]. In our laboratory stimulants are screened for by GC/MS using MSTFA as derivatisating agent after liquid/liquid-extraction at pH 9.2. Unfortunately RA can not be detected by this procedure because MBTFA is necessary for its derivatisation. Moreover, even when MBTFA is used our liquid-liquid extraction at pH 9.2 does not allow to detect RA at the MRPL of 500 ng/ml. Therefore it was decided to include this compound into the LC-MS screening method for diuretics [2].

### **MATERIALS AND METHODS**

#### *Extraction*

Internal standard solution (50 µl mefruside, 20 µg/ml) was added to 2 ml urine, followed by addition of 1 ml sodium acetate buffer (pH 5.2). Liquid-liquid extraction was performed by rolling for 20 min with 4 ml ethyl acetate. After centrifugation the organic layer was transferred into a new tube and evaporated until dry under oxygen free nitrogen at 40 °C. The remaining residue was dissolved in 200 µl mobile phase. 50 µl were injected.

### *Chromatography*

The chromatographic equipment consisted of a Thermo Separation Products (TSP) Model P4000 quaternary pump equipped with a TSP Model AS 3000 autosampler. A Nucleosil C18 column 3 mm x 100 mm, 5  $\mu$ m, protected with a guard column (10 x 2 mm), was used for chromatographic separations (Varian, Sint-Katelijne-Waver, Belgium). The mobile phase was 1% acetic acid (solution A) and acetonitrile. Gradient elution at a flow rate of 0.3 ml/min was as follows: 85% A for 2 min, linear to 45% in 10 min, linear to 35% in 8 min followed by an equilibration step of 8 min at 85% A. Total run time: 28 min.

### *Mass spectrometry*

Detection was performed on a Finnigan LCQ-Deca<sup>®</sup> mass spectrometer.

Ionization of analytes was carried out using Electrospray, positive ionisation (+ESI). The capillary temperature was maintained at 300 °C; the ion source voltage was set at 5000 V and the nebulizer gas was set at 80 units. The capillary voltage was set at 44 V. for the screening full scan MS<sup>2</sup> was performed on the protonated molecular ion [M+H]<sup>+</sup> of ritalinic acid (m/z 220) at a collision energy (CE) of 35, activation Q (Qz): 0.25 and an activation time (AT) of 30 ms.

Full scan MS<sup>3</sup> was performed on the MS<sup>2</sup> product ion m/z 84 (CE: 37, Qz: 0.5, AT: 30 ms).

## **RESULTS AND DISCUSSION**

### *Screening*

RA contains both an amine-function and carboxy-group. Hence protonated as well as deprotonated molecular ions were observed in positive and negative ionization, respectively. Ionization in positive mode was much more abundant and was preferred. Full scan MS<sup>2</sup> generates a single product ion m/z 84 which results from the loss of the benzyl-carboxyl moiety (Figure 1). This ion is monitored in the screening procedure.

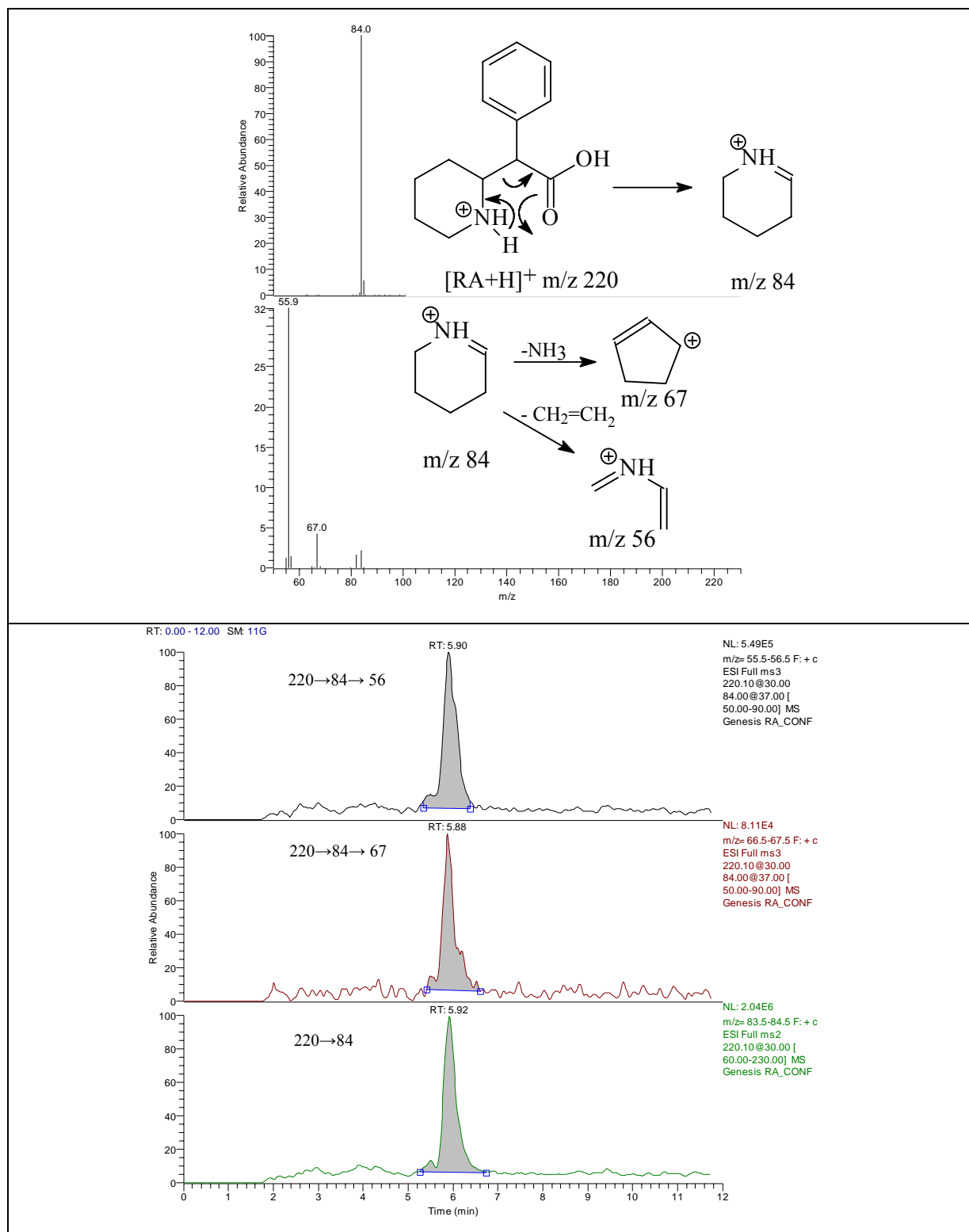


Figure 1: MS<sup>2</sup> and MS<sup>3</sup> spectra for RA (Left). Selected ion chromatograms for RA in a spiked urine sample (500ng/ml) (Right).

### *Confirmation*

For the confirmation 2 additional ions are necessary to fulfill the WADA-criteria concerning reporting an adverse analytical finding [3]. Therefore MS<sup>3</sup> was applied. With the default instrument setting of 0.25 for the Qz-value, the precursor ion m/z 84 could be successfully isolated. Unfortunately no product ions could be detected when CE was applied. When the Qz-value was increased to 0.5 two product ions were detected in MS<sup>3</sup>, m/z 56 and m/z 67 (Figure 1).

According to theoretical principles of ion trap mass spectrometry these product ions should be detectable with the default Qz-value of 0.25 [4]. However several papers [2,5] report the need to adjust the Qz-value to detect low molecular weight fragment ions (mass < m/z 100) as was also required here.

### **CONCLUSION**

Using this LC-MS<sup>2</sup> screening method, the detection of RA can be guaranteed at the MRPL.

For confirmation purposes, the ion trap parameter Qz-value had to be adjusted.

The method has been used routinely since one year and 3 samples were reported as an adverse analytical finding, containing RA. No MPH was detected in these samples.

### **REFERENCES**

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